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LETTERS

# Synthesis of the C-glycosidic analog of adenophostin A, a potent IP<sub>3</sub> receptor agonist, using a temporary silicon-tethered radical coupling reaction as the key step<sup>1</sup>

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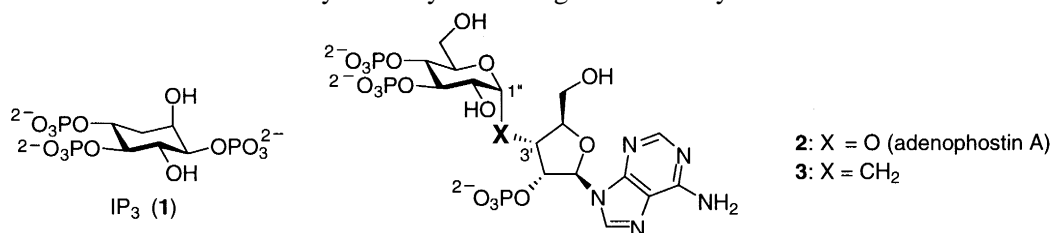
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## Abstract

Synthesis of the C-glycosidic analog (**3**) of adenophostin A, a very potent IP<sub>3</sub> receptor agonist, was achieved using a temporary silicon-tethered reductive radical coupling reaction as the key step. Radical reaction of the silaketal substrate **6** with Bu<sub>3</sub>SnH/AIBN in benzene occurred stereoselectively, and subsequent desilylation gave the desired C-glycosidic disaccharide **7** with the (3 $\alpha$ ,1' $\alpha$ )-configuration as the major product. Compound **7** was converted into the target **3** via the introduction of an adenine base by a Vorbrüggen glycosylation reaction. © 2000 Elsevier Science Ltd. All rights reserved.

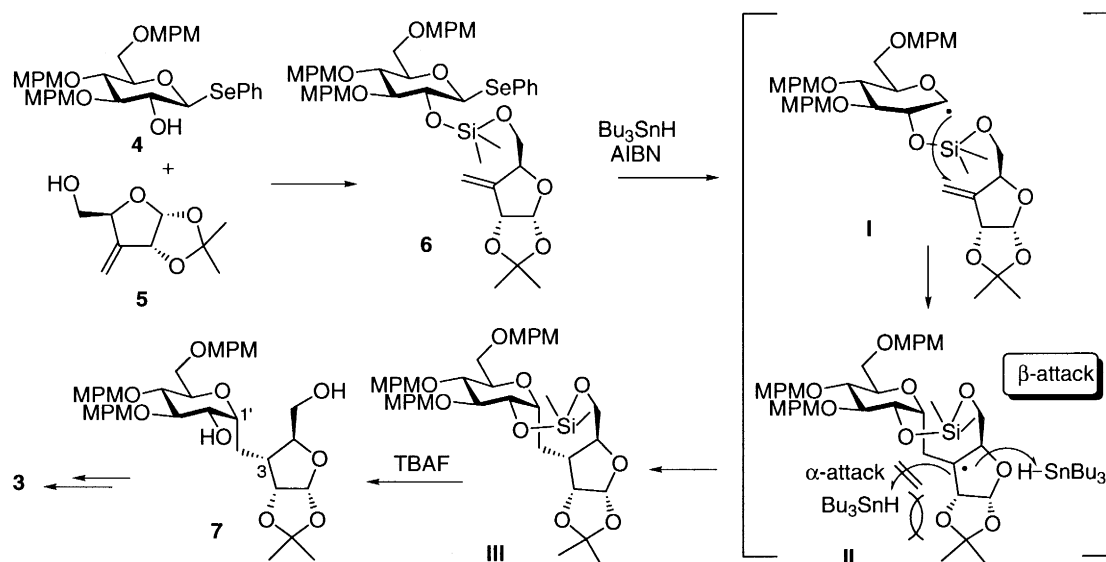
Considerable attention has been focused on D-*myo*-inositol 1,4,5-trisphosphate (IP<sub>3</sub>, **1**), an intracellular Ca<sup>2+</sup>-mobilizing second messenger, because of its biological importance.<sup>2,3</sup> Therefore, much effort has been devoted to the development of specific ligands for the IP<sub>3</sub> receptors, which are very useful for proving the mechanism of IP<sub>3</sub>-mediated Ca<sup>2+</sup> signaling pathways.<sup>4</sup> Recently, adenophostin A (**2**) was isolated from *Penicillium brevicompactum* by Takahashi and co-workers and identified as the strongest IP<sub>3</sub> receptor ligand yet known; **2** is 10–100 times more potent than IP<sub>3</sub> with regard to both the affinity for the IP<sub>3</sub> receptor and the Ca<sup>2+</sup>-mobilizing ability in cells.<sup>5</sup> This finding prompted us to synthesize the C-glycosidic analog **3** of adenophostin A and to examine its biological features,<sup>6</sup> since C-glycosides are known as useful mimics of carbohydrates by enhancing their stability.<sup>7</sup>



In the synthesis of the target compound **3**, the key step is the formation of the C-glycosidic linkage with the desired (3' $\alpha$ ,1'' $\alpha$ )-configuration, as shown in our synthetic plan in Scheme 1. The key C-glycosidic

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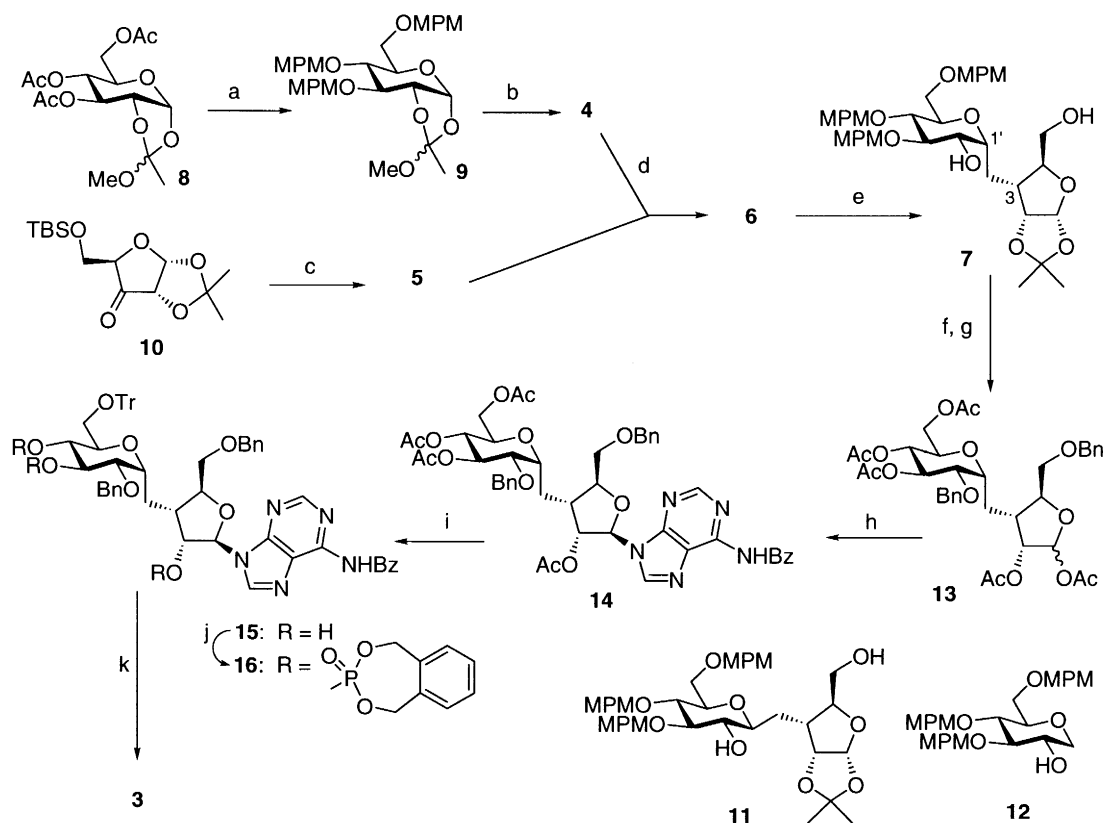
linkage is constructed by a reductive radical coupling reaction of the silaketal tethered<sup>8</sup> substrate **6**, which can be prepared from the pyranose unit **4** and the furanose unit **5**. We assumed that treatment of **6** with Bu<sub>3</sub>SnH/AIBN would produce the 1'-radical **I** in a boat conformation, and that its 1'α-selective cyclization<sup>9</sup> would occur at the sterically unhindered *endo*-position of the 3-methylene to give 3-radical **II**. Subsequent reduction by Bu<sub>3</sub>SnH would occur stereoselectively from the β-face, due to the significant steric repulsion for the isopropylidene group when the reagent approaches the 3-position from the α-face. Accordingly, this radical reaction should proceed stereoselectively to give **III**, and subsequent desilylation would give **7** with the desired (3α,1'α)-configuration. From **7**, the target compound **3** can be synthesized via the introduction of an adenine base by Vorbrüggen's procedure.<sup>10</sup>



Scheme 1.

The synthesis of **3** is summarized in Scheme 2. Removal of the acetyl groups of the known orthoester **8**,<sup>11</sup> which was readily prepared from D-glucose, and subsequent protection of the resulting hydroxyls with *p*-methoxybenzyl (MPM) groups gave **9**. A PhSe group was introduced at the anomeric β-position by treating **9** with PhSeH/MS3Å,<sup>12</sup> and the resulting 2-*O*-acetyl group was removed to complete the synthesis of pyranose unit **4**. On the other hand, a Wittig reaction of a 3-keto sugar **10**,<sup>13</sup> prepared from D-xylose, with Ph<sub>3</sub>P=CH<sub>2</sub> in THF gave the corresponding 3-methylene product, the 5-*O*-TBS group of which was removed with TBAF to give the furanose unit **5**. Next, the units **4** and **5** were temporarily connected with a silaketal linkage. Thus, treatment of **4** with BuLi/Me<sub>2</sub>SiCl<sub>2</sub> in THF yielded the corresponding 2-*O*-Si(Cl)Me<sub>2</sub> product, which was then treated with **5** in the presence of Et<sub>3</sub>N to give the silaketal **6**, the substrate for the radical coupling reaction, in 67% yield.

Next, the reductive coupling reaction of **6** was investigated with Bu<sub>3</sub>SnH/AIBN. When a solution of Bu<sub>3</sub>SnH (2.0 equiv.) and AIBN (0.5 equiv.) in benzene was added slowly over 1.2 h to a solution of **6** in benzene at 80°C, the best results were observed. After the reaction mixture was treated with Bu<sub>4</sub>NF in THF and purified by silica gel flash chromatography, the desired (3α,1'α)-*C*-glycoside **7**<sup>14</sup> was obtained as the major product (50%) along with the *C*-glycoside **11** having the (3α,1'β)-configuration (22%) and the directly reduced product **12** (25%). After protection of the two free hydroxyls of **7** with the benzyl groups, the MPM groups were removed with 90% TFA, and the resulting free hydroxyls were acetylated to give **13**. *N*<sup>6</sup>-Benzoyladenine was successfully introduced at the 1β-position of **13**, using the usual Vorbrüggen glycosylation procedure with a silylated base and SnCl<sub>4</sub> in MeCN to give adenylic *C*-disaccharide **14** in 65% yield. The four acetyl groups of **14** were removed simultaneously, and the 6''-



Scheme 2. Reagents and conditions: (a) (1) NaOMe, THF/MeOH, rt; (2) NaH, MPMCl, HMPA/DMF, rt, 77%; (b) (1) PhSeH, MS3Å, MeNO<sub>2</sub>, reflux; (2) NaOMe, THF/MeOH, rt, 65%; (c) (1) NaOCMe<sub>2</sub>Et, Ph<sub>3</sub>PMeBr, THF, rt; (2) TBAF, THF, rt, 80%; (d) (1) Me<sub>2</sub>SiCl<sub>2</sub>, BuLi, THF, -78°C-rt; (2) **5**, Et<sub>3</sub>N, THF, 0°C-rt, 67%; (e) (1) Bu<sub>3</sub>SnH, AIBN, benzene, reflux; (2) TBAF, THF, 50%; (f) BnBr, NaH, HMPA/DMF/THF, 0°C-rt, 72%; (g) (1) 90% TFA, 0°C-rt; (2) NaOMe, MeOH, rt; (3) Ac<sub>2</sub>O, Et<sub>3</sub>N, DMAP, MeCN, 70%; (h) silylated *N*<sup>6</sup>-benzoyladenine, SnCl<sub>4</sub>, MeCN, 0°C-rt, 65%; (i) (1) NaOMe, MeOH; (2) TrCl, py, 0–50°C, 95%; (j) XEPA, CH<sub>2</sub>Cl<sub>2</sub>, -40°C, then *m*-CPBA, -40°C-rt, 92%; (k) (1) NH<sub>3</sub>, aq. dioxane, rt; (2) H<sub>2</sub>, Pd-black, aqueous MeOH, rt, 89%

primary hydroxyl was selectively protected by a trityl group to give **15**. Phosphate units were introduced, using the phosphoramidite method with *o*-xylene *N,N*-diethylphosphoramidite (XEPA) developed by Watanabe and co-workers.<sup>15</sup> Thus, **15** was treated with XEPA and tetrazole in CH<sub>2</sub>Cl<sub>2</sub>, followed by oxidation with *m*-CPBA to give the desired 2',3'',4''-triphosphate derivative **16** in 92% yield. The *N*<sup>6</sup>-benzoyl group was removed with NH<sub>3</sub>/aq. dioxane. Finally, the trityl- and benzyl-protecting groups were all removed in one step by catalytic hydrogenation with Pd-black in aqueous MeOH to give the target compound **3** in 89% yield as a sodium salt, after treatment with ion-exchange resin.

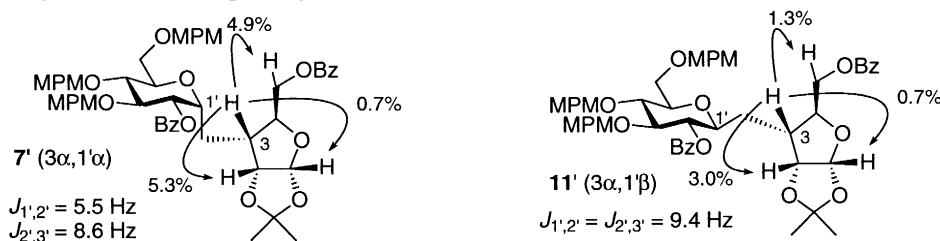
In summary, we have successfully synthesized the *C*-glycosidic analog **3** of adenophostin A, using a temporary silicon-tethered reductive coupling reaction as the key step.<sup>16</sup> Biological evaluation of **3** is under investigation and will be reported elsewhere.

## Acknowledgements

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